Fungal Pneumonia

CASE STUDY





INTRODUCTION **>**

N-chlorotaurine (NCT) is used as a topical antiseptic and anti-infective, and this study aimed to demonstrate the tolerability, safety, and efficacy of inhaled NCT in murine models of fungal pneumonia. Previous methods for organ homogenization post-infection challenge were laborious, so an easier method was established using a FastPrep-24[™] homogenizer and Lysing Matrix M from MP Biomedicals. Survival, viable pathogen load in the lungs and other organs, body weight, organ weight, body temperature, and blood parameters were compared in NCT-treated vs. control groups. Results revealed an improvement of parameters for the NCT-treated *L. corymbifera* pneumonia group and support the safety and tolerability of inhaled NCT for treating fungal pneumonia *in-vivo*.

OVERVIEW **>**

KEYWORDS: N-chlorotaurine (NCT), anti-infective, mouse models, fungal pneumonia, antiseptic, mold, *Lichtheimia, Aspergillus*

AIM OF THE STUDY: Investigate the efficacy of inhaled NCT in a mouse model of fungal pneumonia

APPLICATION: Histology, Blood cell counts, Tissue homogenization, CFU counts

SAMPLE TYPE: Lung, brain, spleen, and kidney tissues from mice

MATERIAL: FastPrep-24[™] 5G homogenizer, Lysing Matrix M • **CASE STUDY:** Efficacy of Inhaled N-Chlorotaurine in a Mouse Model of Lichtheimia corymbifera and Aspergillus fumigatus Pneumonia



5 $\begin{vmatrix} 400 \ \mu L \\ aliquots were removed and diluted in 400 \ \mu L \\ of 0.9\% NaCl for quantitative culture analysis. \end{vmatrix}$



Figure 1. Fungal (a) and bacterial (b) pathogen counts from the homogenized lungs of control (NaCI) and NCT test mice previously challenged with *L. corymbifera*.

CONCLUSION >

Mouse models of fungal pneumonia were used to demonstrate the therapeutic effect of an inhaled antiinfective, NCT, for the treatment of pneumonia. Traditional methods of organ homogenization using a rotating knife or mechanical treatment followed by MixerMill are laborious and time-consuming. A new, easier homogenization method was established using MP Biomedicals' FastPrep-24 5G homogenizer and Lysing Matrix M tubes to process mouse tissues and determine fungal load. The new method proved to be faster and more efficient, allowing for higher sample throughput while also maintaining the viability of fungi and bacteria for culture analysis and determination of microbial load.



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