MagBeads FastDNA® Kit for Blood

Cat. No.: 116574096 (96 PREPS)

Quick-Start Manual

Revision 1.0 Dec 2023

Notes before starting:

- Add 2.5 mL Protease Dissolve Buffer into the Proteinase K, and store at -20 – 8°C after it dissolves.

Manual Extraction

Extraction

Using a microcentrifuge tube

1. Pipette 20 µL Proteinase K and 30 µL Magbeads Particles into the bottom of a 1.5 mL microcentrifuge tube.
2. Add 200 µL sample to the microcentrifuge tube. Use up to 200 µL whole blood, plasma, serum, buffy coat, or body fluids, or up to 5 x 10^6 lymphocytes or culture cells in 200 µL PBS.
3. Add 500 µL Buffer MLA to the sample. Mix by pulse-vortexing for 15 s. Incubate at room temperature for 10 mins with occasional mixing. Vortex occasionally during incubation to disperse the sample, or place it on a shaker.
4. Place the tube on a magnetic stand and allow the beads to aggregate for 1 min. With the tube on the magnetic stand, perform the aspiration, and discard the supernatant from the tube.
5. Add 500 µL Buffer DW1 and resuspend the beads by vortexing for 15 s.
6. Place the tube on a magnetic stand and allow the beads to aggregate for 1 min. With the tube on the magnetic stand, perform the aspiration, and discard the supernatant from the tube.
7. Repeat step 5-6 once more.
8. Add 500 µL Buffer CW and resuspend the beads by vortexing for 20 s.
9. Place the tube on a magnetic stand and allow the beads to aggregate for 1 min. With the tube on the magnetic stand, perform the aspiration, and discard the supernatant from the tube.
10. Leave the tube on the magnetic stand. Slowly add 500 µL Buffer BW3 and leave it for 20-30 s, then aspirate.
11. Add 100 µL Elution Buffer to the sample and shake at maximum speed for 10 mins to elute the DNA. If constant vortexing for 5 mins is not possible, vortex for 20 seconds every 1-2 mins for 10 mins.
12. Place the tube on a magnetic stand and allow the beads to aggregate.
13. Transfer the clear supernatant containing purified DNA into a microcentrifuge tube. Store the DNA at -20°C.

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Extraction

Using a deep-well plate

1. Pipette 20 µL Proteinase K and 30 µL Magbeads Particles into the bottom of a 96 deep-well plate.
2. Add 200 µL sample into the well. Use up to 200 µL whole blood, plasma, serum, buffy coat, or body fluids, or up to 5 x 10^6 lymphocytes or culture cells in 200 µL PBS.
3. Add 500 µL Buffer MLA to the sample. Vortex to mix at shaker (such as IKA MS3) for 10 min at 1000~1200 rpm.
4. Place the plate on a magnetic strand and allow the beads to aggregate for 2 mins. With the plate on the magnetic stand, perform the aspiration, and discard the supernatant from the tube.
5. Add 500 µL Buffer DW1 and vortex to mix at shaker (such as IKA MS3) for 1 min at 1200 rpm.
6. Place the plate on a magnetic stand and allow the beads to separate for 1 min. With the plate on the magnetic stand, perform the aspiration, and discard the supernatant from the tube.
7. Repeat step 5-6 once.
8. Add 500 µL Buffer CW and Vortex to mix at shaker (such as IKA MS3) for 1 min at 1200 rpm.
9. Place the plate on a magnetic stand and allow the beads to separate for 1 min. With the plate on the magnetic stand, perform the aspiration, and discard the supernatant from the tube.
10. Leave the plate on the magnetic stand. Slowly add 500 µL Buffer BW3 and leave it for 30 s, then aspirate.
11. Add 100 µL Elution Buffer to the sample and vortex to mix at shaker (such as IKA MS3) for 10 min at 1300 rpm.
12. Place the plate on a magnetic stand and allow the beads to aggregate. Let it sit at room temperature until the beads are completely cleared from solution.
13. Transfer the clear supernatant containing the purified DNA to a 96-well microplate (not provided). Store the DNA at -20°C

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