MagBeads FastDNA® Kit for FFPE

Cat. No.: 116576096 (96 PREPS)

Quick-Start Manual

Revision 1.0 Dec 2023

Notes before starting:

- Add 2.4 mL Protease Dissolve Buffer into the Proteinase K, and store at -20 - 8℃ after it dissolves.
- Add 1.3 mL Protease Dissolve Buffer to the RNase, and store at -20~8℃ after it dissolves.
- Add 56 mL absolute ethanol to buffer BW1 and store at room temperature

Manual Extraction

Sample Preparation

1. Using a scalpel, trim off the excess paraffin on the sample block. Transfer 1 to 5 sections (10 - 20 µm) of sample into a 1.5 mL microcentrifuge tube.
2. Add 600 µL Buffer DPS into the sample. Vortex for 5 s and briefly centrifuge to bring the sample to the bottom of tube.
3. Incubate at 56°C for 5 mins and vortex vigorously for 15 s to dissolve the paraffin completely.

Note: The Buffer DPS may become opaque or cloudy. If this occurs, add additional Buffer DPS and repeat Step 2.

4. Centrifuge at full speed for 1 min to bring down all FFPE tissues that adhere to the tube wall or underneath the cap. This will create two phases within the solution: an upper dewaxing liquid phase and a lower aqueous phase.

Note: If sample is sufficient, discard the dewaxing liquid to facilitate the operation.

5. Add 200 µL Buffer ATL into the bottom of the tube and add 20 µL Proteinase K into the lower aqueous layer. Mix gently by pipetting up and down.
6. Incubate at 56°C for 60 mins (or until the tissue is completely lysed), followed by 90°C for 60 mins.
7. Briefly centrifuge the tube and transfer the lower aqueous layer into a new microcentrifuge tube and proceed to the extraction step.

Extraction

High Salt Bind

1. Add 20 µL Magbeads Particles N and 400 µL Buffer BST1 to the lysate from step 7 (Sample Preparation). Mix thoroughly by vortex for 10~15 s.
2. Incubate for 3 mins and mix occasionally. Place the tube on the magnetic stand for 2 mins until the beads have formed a tight pellet. Then remove the supernatant.
3. Add 500 µL Buffer BW1 and vortex for 15s to resuspend the beads. Place the tube on the magnetic stand for 2 mins until the beads have formed a tight pellet. Then remove the supernatant.
4. Add 500 µL 75% ethanol, and vortex for 15 s to resuspend the beads. Place the tube on the magnetic stand for 1 min until the beads have formed a tight pellet. Then remove the supernatant.
5. Add 500 µL 75% ethanol, and vortex for 15 s to resuspend the beads. Place the tube on the magnetic stand for 1 min until the beads have formed a tight pellet. Then remove the supernatant.
6. Centrifuge briefly to collect the liquid on the tube and remove all the liquid. Air dry for 10 mins.
7. Add 30~100 µL Elution Buffer to the sample, re-suspend the beads by vortex. Incubate at 55°C for 10 mins with shaking. If there is no shaking device, vortex 2~3 times to mix.
8. Place the tube on the magnetic rack for 3 mins. Transfer the supernatant containing the purified DNA to a clean 1.5 mL centrifuge tube.

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Extraction

Ethanol Bind

1. Add 20 µL Magbeads Particles N and 200 µL Buffer BST1 to the lysate from step 7 (Sample Preparation). Mix thoroughly by vortex from 10 s.
2. Add 300 µL absolute ethanol to the sample, mix thoroughly by inverting for 15~30 times. Incubate for 3 mins and mix occasionally. Place the tube on the magnetic stand for 1 min until the beads have formed a tight pellet. Then remove the supernatant.
3. Add 500 µL Buffer BW1 and vortex for 15 s to resuspend the beads. Place the tube on the magnetic stand for 2 mins until the beads have formed a tight pellet. Then remove the supernatant.
4. Add 500 µL 75% ethanol, and vortex for 15 s to resuspend the beads. Place the tube on the magnetic stand for 1 min until the beads have formed a tight pellet. Then remove the supernatant.
5. Add 500 µL 75% ethanol, and vortex for 15 s to resuspend the beads. Place the tube on the magnetic stand for 1 min until the beads have formed a tight pellet. Then remove the supernatant.
6. Centrifuge briefly to collect the liquid on the tube and remove all the liquid. Air dry for 10 mins.
7. Add 30~100 µL Elution Buffer to the sample, re-suspend the beads by vortex. Incubate at 55°C for 10 mins with shaking. If there is no shaking device, vortex 2~3 times to mix.
8. Place the tube on the magnetic rack for 3 mins. Transfer the supernatant containing the purified DNA to a clean 1.5 mL centrifuge tube.

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