# SPINeasy™ DNA Kit for Yeast

Cat. No.: 116557050 ( 50 PREPS ) / 116557000 ( 5 PREPS )

## Quick-Start Protocol

Revision Oct 2023

Scan QR code for more information from instruction manual

**Notes before starting:**

- This protocol requires a centrifuge capable of generating at least 15,000 *g* to obtain optimal results.
- **Wash Buffer Y2** is supplied as a concentrate. Before using for the first time, add absolute alcohol as indicated on the bottle to obtain a working solution.

### Column preparation

- Add **200 µL Equilibration Buffer** into the **Column S** placed in **Collection tube** (provided), centrifuge for **1 min @ 15,000 *g***. Discard the flow-through and reuse Collection tube.

### Sample preparation

- Harvesting cells from liquid culture

  Pellet the cells 1-2 mL (maximum **4 x $10^8$ cells**) by centrifuging for **5 min @ 15,000 *g***. Discard the supernatant.
- Harvesting colonies from solid medium

  Scrape a single colony (**≥ 2 mm** in diameter) or amount **< 50 mg** from solid medium.

### Lysis

1. Add **5 µL RNase A** and **560 µL Lysis Buffer Y** to each microcentrifuge tube of pellet collected. Re-suspend the pellet thoroughly by pipetting repeatedly. Transfer all lysate to **Lysing Matrix YG** tube.
2. Homogenize in a FastPrep® Instrument for **35 sec** at speed setting of **5.0 m/sec** or vortex for **10 min** at speed setting of **2,500 rpm**.
3. Centrifuge for **2 min @ 15,000 *g***.

   ***Note:*** *Cells* **> 4 x$10^8$** *or colonies* **≥ 50 mg**, *homogenize twice.*

### Bind

4. Slowly add **560 µL Binding Buffer Y** into the Lysing Matrix YG tube, then mix thoroughly.
5. Centrifuge for **1 min @ 15,000 *g***.
6. Carefully transfer **750 µL** of the supernatant to Column S.
7. Centrifuge for **1 min @ 15,000 *g***.

### Wash

8. Add **600 µL Wash Buffer Y1** to the wall of Column S. Centrifuge for **1 min @ 15,000 *g***. Discard the flow-through.
9. Add **700 µL Wash Buffer Y2** to the wall of Column S. Centrifuge for **1 min @ 15,000 *g***. Discard the flow-through.
10. Centrifuge Column S for **2 min @ 15,000 *g*** to dry the column.

### Elute

11. Transfer the column into a clean **Elution tube** (provided). Add **50-200 µL Elution Buffer EB** or ultrapure water **(pH >7.0)** (not provided) to the center of column, wait for **2 min**, and centrifuge for **1 min @ 15,000 *g***. The DNA samples are now ready for downstream application.

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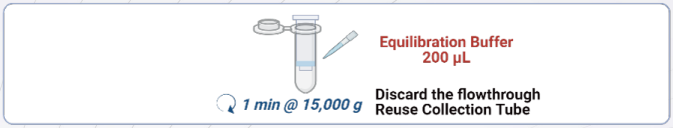
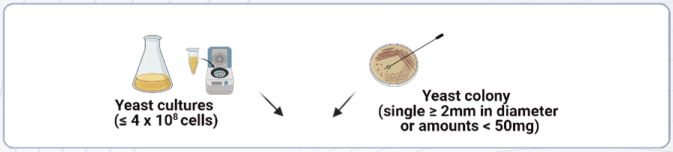
Flow-Chart

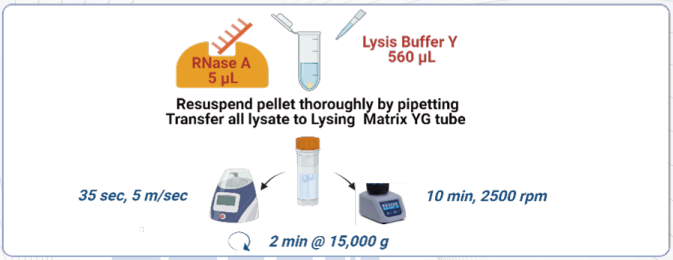
### Column preparation



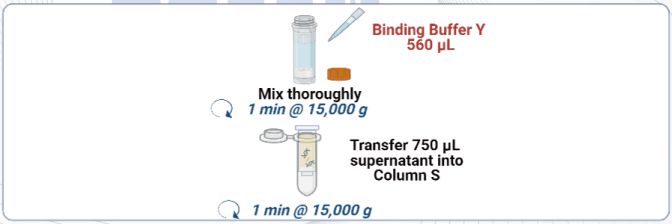

### Sample preparation



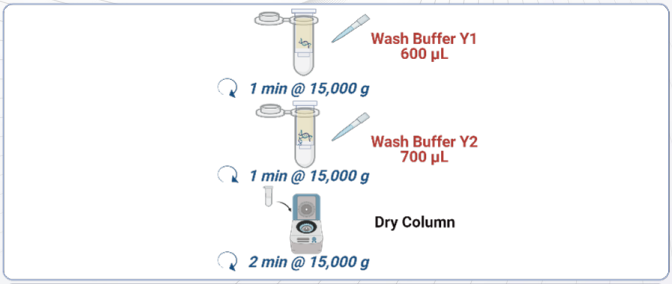

### Lysis



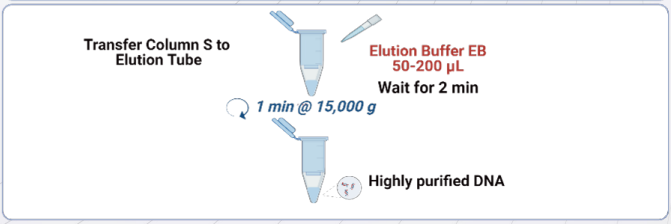

### Bind




### Wash




### Elute





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